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Origination and establishment of a trigenic reproductive isolation system in rice

Yidan Ouyang, Guangwei Li, Jiaming Mi, Conghao Xu, Hongyi Du, Chengjun Zhang, Weibo Xie, Xianghua Li, Jinghua Xiao, Huazhi Song, Qifa Zhang



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| 1 | Origination and establishment of a trigenic reproductive isolation system in rice |
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| 3 | Yidan Ouyang ^{1†*} , Guangwei Li ^{1†} , Jiaming Mi ^{1†} , Conghao Xu ¹ , Hongyi Du ¹ , Chengjun |
| 4 | Zhang ² , Weibo Xie ¹ , Xianghua Li ¹ , Jinghua Xiao ¹ , Huazhi Song ¹ , Qifa Zhang ¹ |
| 5 | |
| 6 | ¹ National Key Laboratory of Crop Genetic Improvement and National Centre of Plant |
| 7 | Gene Research (Wuhan), Huazhong Agricultural University, Wuhan 430070, China; |
| 8 | ² Germplasm Bank of Wild Species, Kunming Institute of Botany, Chinese Academy |
| 9 | of Sciences, No. 132, Lanhei Road, Kunming 650201, Yunnan, China |
| 10 | |
| 11 | [†] These authors contributed equally to this work. |
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| 13 | *Author for correspondence: |
| 14 | Yidan Ouyang |
| 15 | Tel: +86 27 87281677 |
| 16 | Fax: +86 27 87287092 |
| 17 | Email: diana1983941@mail.hzau.edu.cn |
| 18 | |
| 19 | Running title: Origin of a reproductive isolation system in rice |

| 20 | Dear Editor, |
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| 21 | Reproductive isolation is both the indicator and a primary force of speciation, and |
| 22 | plays a key role in maintaining species identity. Understanding the origin and |
| 23 | mechanisms of reproductive isolation is of fundamental importance in evolutionary |
| 24 | biology. In recent years, a number of genes that induce reproductive barriers have |
| 25 | been identified in several model organisms such as Drosophila, rodents, yeast, |
| 26 | Arabidopsis, rice and other species, which have greatly advanced our understanding |
| 27 | of the mechanisms regulating reproductive isolation (Chae et al., 2014; Lafon-Placette |
| 28 | and Kohler, 2015; Maheshwari and Barbash, 2011; Ouyang and Zhang, 2013 review |
| 29 | thereof). However, little is known about how the genomes evolve to form |
| 30 | reproductive barriers and how such systems become established in the populations, |
| 31 | although such reproductive barriers are widely observed and demand understanding |
| 32 | for both biological study of speciation and practical application in crop improvement. |
| 33 | The Asian cultivated rice (Oryza sativa L.) comprises two subspecies, indica and |
| 34 | japonica. Hybrid sterility between indica and japonica represents one of the |
| 35 | best-characterized examples of postzygotic reproductive isolation in plants. We |
| 36 | previously reported a killer-protector system at the S5 locus composed of three |
| 37 | tightly-linked genes, ORF3, ORF4, and ORF5, which together regulated the female |
| 38 | gamete fertility of indica-japonica hybrids (Chen et al., 2008; Yang et al., 2012). |
| 39 | Typical <i>indica</i> and <i>japonica</i> varieties contain the haplotypes of <i>ORF3+ORF4-ORF5+</i> |
| 40 | and ORF3-ORF4+ORF5-, respectively. The killer is made of ORF4+ and ORF5+, |
| 41 | which work together to kill the gamete, while ORF3+ confers protection to the |
| 42 | gametes, such that female gametes without ORF3+ are selectively eliminated, causing |
| 43 | hybrid sterility and segregation distortion. |
| 44 | Reciprocal BLASTN search using ORF3, ORF4, and ORF5 as queries identified |
| 45 | a unique set of three tightly-linked genes on chromosome 5 (designated as Ospara3-5) |
| 46 | showing 73.72%, 67.74%, and 71.37% identities to ORFs3, 4, 5 (chromosome 6), |
| 47 | respectively (Supplemental Table S1). TBLASTN search identified another set of |
| 48 | three genes locating adjacent to each other on chromosome 12 (designated as |
| 49 | Os345-like), which showed less similarity to either ORFs3-5 or Ospara3-5. |

| 50 | Sequences with significant similarity to either ORFs3-5, Ospara3-5 or Os345-like |
|----------|---|
| 51 | were identified in other species by reciprocal blast (Supplemental Table S1). These |
| 52 | similar sequences also located adjacent to each other, although they have frequently |
| 53 | changed their relative orientations. One group of similar sequences showed higher |
| 54 | similarity with ORFs3-5 or Ospara3-5 (labeled as para), and the other group was |
| 55 | more similar to Os345-like (labeled as like). Therefore, ORF3-5/Ospara3-5 and |
| 56 | Os345-like belonged to two different lineages, which led us to focus our attention to |
| 57 | the ORF3-5/Ospara3-5 pair relationship in subsequent pursuit for the origin of S5. |
| 58 | Based on phylogenetic trees, the Ospara3-5 were more closely related to most hits in |
| 59 | the outgroup species than to ORFs3-5 (Supplemental Figures S1A, B, C). Thus the |
| 60 | Ospara3-5 are likely to be more ancestral than the S5 locus, suggesting that ORFs3-5 |
| 61 | were derived from Ospara3-5. Interestingly, we found a TBLASTN hit in Z. latifolia |
| 62 | (ZlORF4), which belonged to the Oryzeae tribe, showed higher similarity to ORF4 |
| 63 | than to other sequences (sequences labeled as para). Therefore, the three-gene |
| 64 | fragment of the S5 complex might have originated from Ospara3-5 after the |
| 65 | formation of the Oryzeae tribe (Supplemental Figure S2). |
| 66 | A question thus arose: how did the Ospara3-5 fragment duplicate to form the |
| 67 | ORFs3-5 block? We investigated the sequence similarity between the ORFs3-5 and |
| 68 | Ospara3-5 fragments, which identified several intergenic regions showing high |
| 69 | similarity between the two homologous blocks (Figure 1A, Supplemental Table S2). |
| 70 | These regions contained multiple repetitive sequences of the RPO_OS Helitron |
| 71 | transposon elements (http://www.girinst.org/) (Kapitonov and Jurka, 2007). We |
| 72 | detected Helitron termini bracketing Ospara4 and Ospara5 (Hel_Os05g-A). And |
| 73 | similarly, ORF4 and ORF5 were also flanked by Helitron termini (Hel_Os06g-A). |
| 74 | This suggested a possibility that the ORFs3-5 segment was derived by Helitron |
| 75 | movement, carrying the internal Ospara4-5 fragment on chromosome 5 and capturing |
| 76 | |
| | the 3'-terminal Ospara3, to insert them on chromosome 6 thus generating a new |
| 77 | the 3'-terminal <i>Ospara3</i> , to insert them on chromosome 6 thus generating a new fragment containing <i>ORFs3</i> –5 (Supplemental Figure S3). This inference was partly |
| 77 78 | |

| 80 | and to capture the DNA fragment downstream (Kapitonov and Jurka, 2007). After |
|-----|---|
| 81 | duplication through <i>Helitron</i> transposition, a combination of mutational mechanisms |
| 82 | acted together to recreate the three genes of the S5 locus from Ospara3-5 |
| 83 | (Supplemental Figures S4A, B, C), thus conferred functional divergence to the |
| 84 | newborns giving rise to reproductive barriers. |
| 85 | To learn how the three genes evolved into a reproductive barrier, we analyzed |
| 86 | patterns of variation and diversity of ORFs3-5 in 635 rice accessions from 62 |
| 87 | geographically diverse countries, representing two cultivated and 11 wild species |
| 88 | (Supplemental Table S3, S4). The ORF3+ORF4+ORF5+ was the most frequent type |
| 89 | (56.94%) in the wild species (Supplemental Table S5), and ORF3+, ORF4+, and |
| 90 | ORF5+ were the most frequent alleles in both O. sativa (0.6735, 0.4822, and 0.5797) |
| 91 | and wild species (0.9306, 0.7222, and 0.7639) (Supplemental Table S6), suggesting |
| 92 | that they may be either the ancestral alleles or under selection. We further analyzed |
| 93 | the functional Indels/SNPs of ORFs3-5 in their parent genes. Interestingly, the 11-bp |
| 94 | sequence characteristic of ORF4+ and functional SNP of C819 characteristic of |
| 95 | ORF5+ were identified in Ospara4 and Ospara5, respectively, suggesting that they |
| 96 | were the ancestral alleles (Supplemental Figure S4D) (Yang et al., 2012). |
| 97 | The ancestral ORF3+ORF4+ORF5+ represented a balance between killing and |
| 98 | protecting. Therefore, mutation in the protector ORF3+ alone would be unable to |
| 99 | survive because of a functional killer. Thus the mutation of ORF3+ to ORF3- must |
| 100 | have occurred after the emergence of non-functional killer, either ORF4- or ORF5- |
| 101 | (including ORF5n). Based on our data, a mutation that occurred in ORF4+ in the |
| 102 | ancestral populations resulted in ORF3+ORF4-ORF5+ giving rise to the typical |
| 103 | indica rice. Similarly, ORF5- emerged in the ancient population with |
| 104 | ORF3+ORF4+ORF5+ to generate the intermediate population carrying |
| 105 | ORF3+ORF4+ORF5 An important question was whether ORF3- arose in the |
| 106 | population carrying ORF3+ORF4-ORF5+ or the one with ORF3+ORF4+ORF5 |
| 107 | The absence of ORF3-ORF4-ORF5+ ruled out the first possibility. In contrast, |
| 108 | ORF3-ORF4+ORF5- was observed in high frequency indicating that the ORF3- allele |
| 109 | arose in the population carrying ORF3+ORF4+ORF5-, thus generating the typical |

| 110 | japonica genotype ORF3-ORF4+ORF5- and subsequently spread in the population |
|-----|--|
| 111 | (Supplemental Figure S5). The inference was well supported by the phylogenetic tree |
| 112 | (Supplemental Figure S6) and the co-existence of ORF3+ORF4+ORF5- and |
| 113 | ORF3-ORF4+ORF5- in wild rice accessions from Jiangxi Province of China |
| 114 | (Supplemental Table S3). Consequently, reproductive barrier occurred in |
| 115 | hybridization between the populations carrying ORF3+ORF4-ORF5+ and ones with |
| 116 | ORF3-ORF4+ORF5 |
| 117 | An interesting question now is: what are the evolutionary forces that drive the |
| 118 | mutually incompatible haplotypes, ORF3+ORF4-ORF5+ and ORF3-ORF4+ORF5-, |
| 119 | to high frequencies to establish such at the population level? We investigated variation |
| 120 | patterns of 4000 SNPs in regions flanking S5 (~154-kb) in 529 accessions of O. |
| 121 | sativa (http://ricevarmap.ncpgr.cn/django/home/) (Figure 1B). A dramatic decrease in |
| 122 | genetic variation was observed around S5 region (~44kb) in the indica subgroup, |
| 123 | which is in contrast to the patterns observed in other rice groups especially compared |
| 124 | to the temperate japonica subgroup. We further conducted tests for selection in |
| 125 | different rice lineages (Supplemental Table S7). Significant negative values were |
| 126 | detected for all the three genes at S5 locus in indica rice using Fay and Wu's H, Fu |
| 127 | and Li's D* and F* tests, suggesting that the spread of the <i>indica</i> haplotype might be |
| 128 | driven by natural selection, possibly due to fitness advantage. We also determined |
| 129 | whether demography is responsible for the departure from neutrality of S5 locus in |
| 130 | indica rice. The coalescent simulation results revealed significant values for Fay and |
| 131 | Wu's H, Fu and Li's D* and F* tests in <i>indica</i> populations for all the three genes at S5 |
| 132 | locus (Supplemental Table S8). Taken together, the results suggested that the indica |
| 133 | haplotype of S5 locus was under strong positive selection during the evolution and/or |
| 134 | rice breeding. Such positive selection may have provided a driving force leading to |
| 135 | the establishment of <i>ORF3+ORF4-ORF5+</i> to dominate the <i>indica</i> group. |
| 136 | By contrast, the <i>japonica</i> haplotype ORF3-ORF4+ORF5- also reached a high |
| 137 | frequency after origination within a short period of time. However, using the allele |
| 138 | frequency spectrum tests and demographic simulations, not all the tests returned |
| 139 | significant values for the three genes, suggesting that selection might not be |

| 140 | responsible for the spread of the japonica haplotype. Therefore, the preservation of |
|--|---|
| 141 | typical japonica genotype (ORF3-ORF4+ORF5-) might be due to the founder effect |
| 142 | resulting from domestication of the <i>japonica</i> rice. This inference was highly plausible |
| 143 | when considering that japonica subspecies had experienced a severe bottleneck |
| 144 | during domestication of rice (Huang et al., 2012; Zhu et al., 2007). |
| 145 | In summary (Supplemental Figure S7), the S5 complex originated by duplication |
| 146 | from Ospara3-5 after the formation of the Oryzeae tribe, most likely through Helitron |
| 147 | transposition. A combination of mutational steps generated incompatible indica and |
| 148 | japonica alleles in pre-differentiated rice groups, giving rise to the trigenic |
| 149 | reproductive isolation system. Natural selection in indica rice and founder effect |
| 150 | associated with domestication in japonica populations increased the frequencies of |
| 151 | incompatible alleles to form a functional reproductive barrier between the indica and |
| 152 | japonica subspecies, eventually result in genetic differentiation and restructuring of |
| 153 | rice genetic composition. This also suggests the possibility that domestication and |
| 154 | artificial breeding in crops can lead to the early stages of speciation. |
| 155 | |
| 156 | ACCESSION NUMBERS |
| 157 | XXXX to XXXX |
| 158 | |
| | |
| 159 | AUTHOR CONTRIBUTIONS |
| | AUTHOR CONTRIBUTIONS YO and QZ conceived and designed the experiments; YO, GL, JM, CX, and HD |
| 159 | |
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| 159 160 161 | YO and QZ conceived and designed the experiments; YO, GL, JM, CX, and HD performed the experiments, including the PCR amplification, DNA sequencing, and |
| 159 160 161 162 | YO and QZ conceived and designed the experiments; YO, GL, JM, CX, and HD performed the experiments, including the PCR amplification, DNA sequencing, and sequence assembling; YO, GL, JM, CX, and CZ analyzed the data; WX, XL, JX, and |
| 159 160 161 162 163 | YO and QZ conceived and designed the experiments; YO, GL, JM, CX, and HD performed the experiments, including the PCR amplification, DNA sequencing, and sequence assembling; YO, GL, JM, CX, and CZ analyzed the data; WX, XL, JX, and |
| 159 160 161 162 163 164 | YO and QZ conceived and designed the experiments; YO, GL, JM, CX, and HD performed the experiments, including the PCR amplification, DNA sequencing, and sequence assembling; YO, GL, JM, CX, and CZ analyzed the data; WX, XL, JX, and HS contributed reagents/materials/analysis tools; YO and QZ wrote the paper. |
| 159 160 161 162 163 164 165 | YO and QZ conceived and designed the experiments; YO, GL, JM, CX, and HD performed the experiments, including the PCR amplification, DNA sequencing, and sequence assembling; YO, GL, JM, CX, and CZ analyzed the data; WX, XL, JX, and HS contributed reagents/materials/analysis tools; YO and QZ wrote the paper. ACKNOWLEDGMENTS |
| 159 160 161 162 163 164 165 166 | YO and QZ conceived and designed the experiments; YO, GL, JM, CX, and HD performed the experiments, including the PCR amplification, DNA sequencing, and sequence assembling; YO, GL, JM, CX, and CZ analyzed the data; WX, XL, JX, and HS contributed reagents/materials/analysis tools; YO and QZ wrote the paper. ACKNOWLEDGMENTS We are indebted to Drs. Manyuan Long, Hanhui Kuang, Li Zhang, Bin He, Xiaoming |

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| 177 | |
| 178 | CONFLICT OF INTEREST |
| 179 | All the authors declare that they have no conflict of interests. |
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| 182 | REFERENCES |
|-----|---|
| 183 | Chae, E., Bomblies, K., Kim, S.T., Karelina, D., Zaidem, M., Ossowski, S. |
| 184 | Martin-Pizarro, C., Laitinen, R.A., Rowan, B.A., Tenenboim, H., et al. (2014). |
| 185 | Species-wide genetic incompatibility analysis identifies immune genes as hor |
| 186 | spots of deleterious epistasis. Cell 159:1341-1351. |
| 187 | Chen, J., Ding, J., Ouyang, Y., Du, H., Yang, J., Cheng, K., Zhao, J., Qiu, S., Zhang, |
| 188 | X., Yao, J., et al. (2008). A triallelic system of S5 is a major regulator of the |
| 189 | reproductive barrier and compatibility of indica-japonica hybrids in rice. Proc |
| 190 | Natl Acad. Sci. USA 105:11436-11441. |
| 191 | Huang, X., Kurata, N., Wei, X., Wang, Z.X., Wang, A., Zhao, Q., Zhao, Y., Liu, K., Lu |
| 192 | H., Li, W., et al. (2012). A map of rice genome variation reveals the origin of |
| 193 | cultivated rice. Nature 490:497-501. |
| 194 | Kapitonov, V.V., and Jurka, J. (2007). Helitrons on a roll: eukaryotic rolling-circle |
| 195 | transposons. Trends Genet. 23:521-529. |
| 196 | Lafon-Placette, C., and Kohler, C. (2015). Epigenetic mechanisms of postzygotic |
| 197 | reproductive isolation in plants. Curr. Opin. Plant Biol. 23:39-44. |
| 198 | Maheshwari, S., and Barbash, D.A. (2011). The genetics of hybrid incompatibilities |
| 199 | Annu. Rev. Genet. 45:331-355. |
| 200 | Ouyang, Y., and Zhang, Q. (2013). Understanding reproductive isolation based on the |
| 201 | rice model. Annu. Rev. Plant Biol. 64:111-135. |
| 202 | Yang, J., Zhao, X., Cheng, K., Du, H., Ouyang, Y., Chen, J., Qiu, S., Huang, J., Jiang. |
| 203 | Y., Jiang, L., et al. (2012). A killer-protector system regulates both hybrid |
| 204 | sterility and segregation distortion in rice. Science 337:1336-1340. |
| 205 | Zhu, Q., Zheng, X., Luo, J., Gaut, B.S., and Ge, S. (2007). Multilocus analysis of |
| 206 | nucleotide variation of Oryza sativa and its wild relatives: severe bottleneck |
| 207 | during domestication of rice. Mol. Biol. Evol. 24:875-888. |
| 208 | |
| 200 | |

FIGURE LEGENDS

| Figure 1. Sequence (A) and genetic variation (B) analysis in S5 region. Sequence |
|---|
| analysis is performed between S5 and its paralogous regions in rice. The numbers in |
| brown and black indicate genomic positions on the chromosomes (in Mb) and the |
| relative scales (kb) in the analyzed regions, respectively. Nucleotide diversity is |
| calculated based on polymorphism data flanking S5 locus (4000 SNPs) in 529 rice |
| accessions of O. sativa. The position of the S5 region is shown at the bottom. See |
| methods for details. |

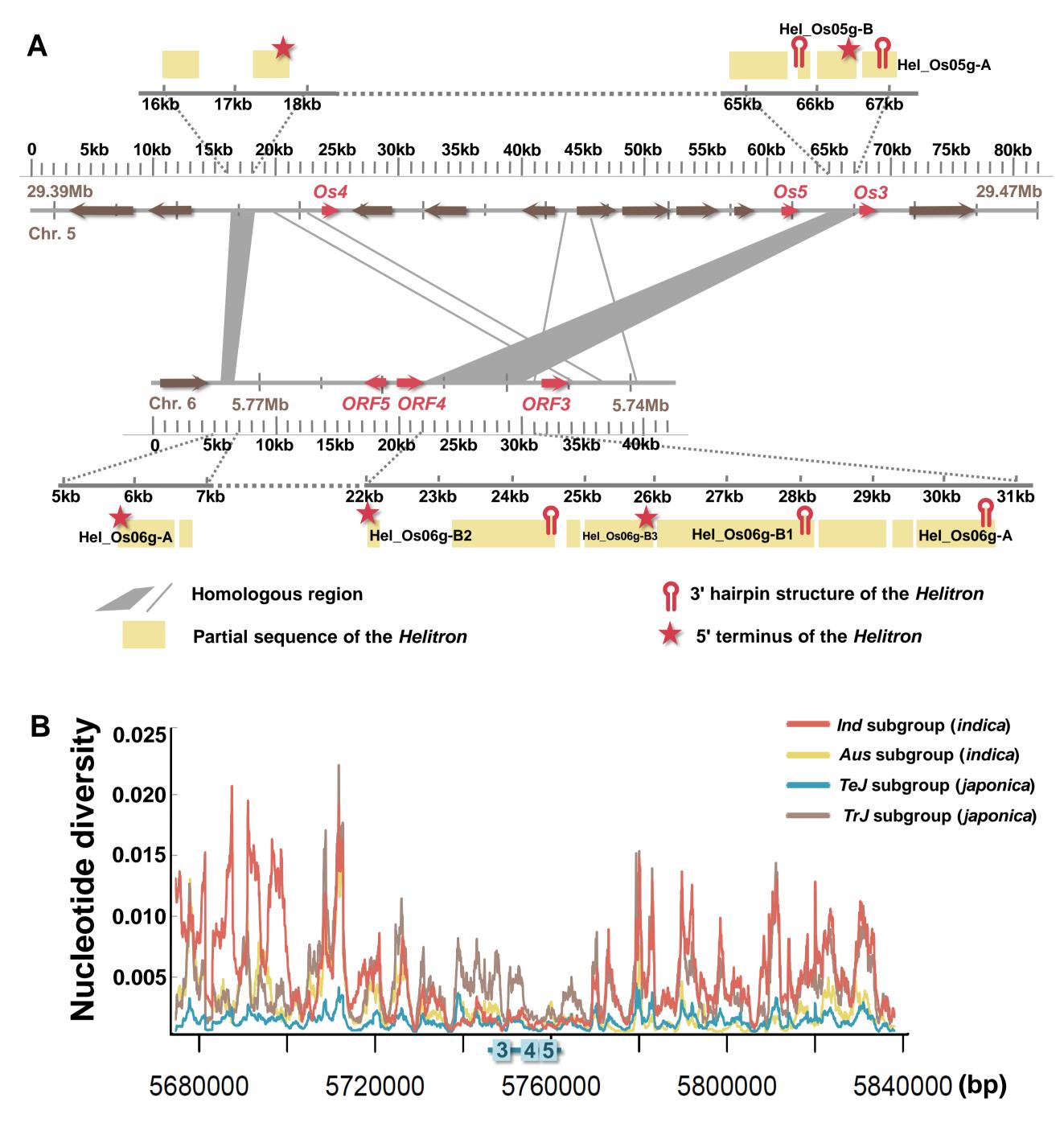


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